

SAXITOXIN ELISA

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A competitive enzyme immunoassay for
screening and quantitative analysis of
Saxitoxin in various matrices

EUROPROXIMA SAXITOXIN ELISA

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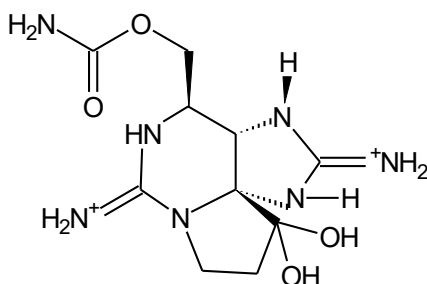
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BRIEF INFORMATION

The saxitoxin ELISA is a competitive enzyme immunoassay for the screening and quantitative analysis of the shellfish poison saxitoxin in various matrices. The test is based on rabbit polyclonal antibodies against saxitoxin. With this ELISA, 96 analyses can be performed. Samples and standards are measured in duplicate which means that in total 40 samples can be analysed. The ELISA kit contains all reagents required to perform the test, including ready-to-use standards. Materials and chemicals necessary for the extraction of saxitoxin from sample material are not included in the test kit

1. INTRODUCTION



Chemical structure of saxitoxin (STX)

Marine biotoxins, also called phytotoxins or shellfish toxins, are naturally occurring compounds produced by algae and phytoplankton. Under normal conditions these compounds do not cause any problem. However, filter feeders such as clams, mussels, oysters and crustaceans can consume large quantities of these algae when environmental conditions result in harmful algal blooms, also known as 'Red Tides'. High concentrations of shellfish toxins then accumulate in these animals causing illness amongst people who eat them. There are four syndromes called shellfish poisoning, i.e. paralytic shellfish poisoning (PSP), neurotoxic shellfish poisoning (NSP), diarrhetic shellfish poisoning (DSP) and amnesic shellfish poisoning (ASP). A variety of symptoms are reported to arise from these poisonings, including stomach cramps, abdominal pain, diarrhea, headaches, memory loss, paralysis and in some cases even death.

PSP is caused by saxitoxin and its analogues. DSP is primarily caused by okadaic acid (OA) and several analogues of OA, whereas ASP is caused by domoic acid (DA).

Saxitoxin (STX) is one of the most potent natural toxins known. It acts as a selective sodium channel blocker and is produced by certain species of marine dinoflagellates (*Alexandrium* sp., *Gymnodinium* sp., *Pyrodinium* sp.) and cyanobacteria (*Anabaena* sp., some *Aphanizomenon* spp., *Cylindrospermopsis* sp., *Lyngbya* sp., and *Planktothrix* sp.). The term saxitoxin can also refer to the entire suite of related neurotoxin produced by these microorganisms, such as pure saxitoxin (STX), neosaxitoxin (neoSTX), decarbamoylsaxitoxin (dcSTX) and the gonyautoxins (GTX).

In the European Union, Regulation (EC) no 853/2004 stipulates that live bivalve mollusks must not contain Paralytic Shellfish Poisoning toxins (saxitoxin and analogues) in total quantity (measured in the whole body or any part edible separately) that exceeds a limit of 800 µg per kilogram.

2. PRINCIPLE OF THE SAXITOXIN ELISA

The microtiter plate-based ELISA kit consists of 12 strips, each containing 8 wells, precoated with sheep antibodies to rabbit IgG. Antibodies (rabbit polyclonal anti-saxitoxin antibodies), horseradish peroxidase (HRP) labelled saxitoxin (enzyme conjugated saxitoxin-HRP) as well as saxitoxin standard solutions or samples are added to the precoated wells, followed by a single incubation step. The rabbit anti-saxitoxin antibodies are bound by the immobilized sheep anti-rabbit antibodies and simultaneously the saxitoxin-HRP and saxitoxin present in the standard solution or sample compete for binding to the anti-saxitoxin antibody (competitive enzyme immunoassay). After incubation for 30 minutes, non-bound reagents are removed in a washing step.

The amount of bound saxitoxin-HRP is visualised by the addition of a substrate/chromogen (H_2O_2 /TMB; TetraMethylBenzidine). The colourless chromogen is converted by the enzyme (HRP) into a blue reaction product. The more saxitoxin present in the standard solution or sample, the less colour is developed. The substrate is stopped by the addition of sulfuric acid. In the acidic environment the blue colour changes into a yellow colour. The obtained colour intensity is measured photometrically at 450 nm and is inversely proportional to the saxitoxin concentration in the sample.

3. SPECIFICITY AND SENSITIVITY

For this saxitoxin ELISA a specific antibody is used, obtained by immunisation of rabbits with a saxitoxin protein conjugate. The reactivity pattern of the antibody as tested in buffer is:

Saxitoxin	100%
Decarbamoyl saxitoxin	19.2%
Gonyautoxins, GTX 2/3	5.6%
GTX 5	26.2%
Decarbamoyl GTX 2/3	0.2%
Neosaxitoxin	1.4%
Decarbamoyl neosaxitoxin	0.5%
Gonyautoxins GTX 1/4	< 0.1%
C 1/2	0.2%

The cross-reactivities are determined in a buffer system. The reported values may be different in samples due to matrix effects.

The test cannot discriminate between analytes and cross-reactive substances.

The Limit of detection (LOD is determined under optimal conditions. Cut-off criteria need critical consideration.

Matrix	LOD (ppb)
Mussel	10
Oyster	5

If the sample is found to be non-compliant, the results shall be verified by re-analysis of the sample using a confirmatory method.

4. HANDLING AND STORAGE

- Saxitoxin is subject to degradation when exposed to the light or warmth.
- Kit and kit components should be stored at 2°C to 8°C in a dark place. For repeated use store kit components as specified under chapter 9.
- After the expiry date of the kit and/or components has passed, no further quality guarantee is valid.
- Bring all kit components including the microtiter plate to ambient (room) temperature before use.
- Dilute the kit components immediately before use, but after the components are brought to ambient temperature.
- Avoid condensation in the wells of the plate. Bring the sealed plate to ambient temperature before opening the plate sealing.
- The substrate chromogen solution can be stored in a refrigerator (2°C to 8°C) until the expiry date stated on the label.
- Exposure of the chromogen solution to light should be avoided.

Degeneration of the reagents may have occurred when the following phenomena are observed:

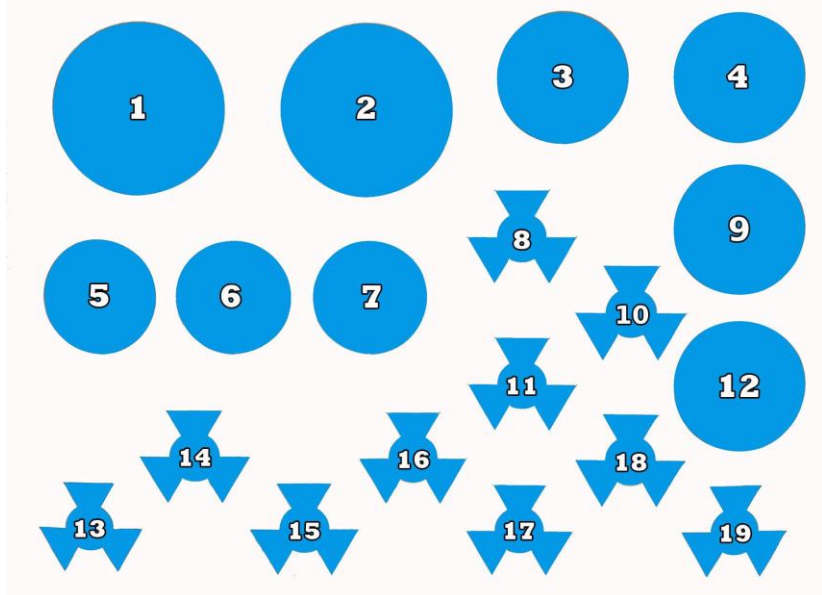
- A blue colouring of the chromogen solution before transferring it into the wells.
- A weak or no colour reaction in the zero standard wells ($E_{450nm} < 0.8$).

5. KIT CONTENTS

Manual

1 sealed microtiter plate (12 strips, 8 wells each), coated with purified sheep anti-rabbit IgG

Position of the reagents in the kit. For preparation of the reagents see Chapter 9.



1. **Dilution buffer** (30 ml, 10x concentrated)
2. **Rinsing Buffer** (30 ml, 20x concentrated)
3. **Substrate solution** (12 ml, Ready-to-use)
4. **Stop solution** (6 ml, Ready-to-use)
5. **Conjugate** (lyophilised, blue cap)
6. not in use
7. not in use
8. not in use
9. not in use
10. **Antibody** (100x concentrated, yellow cap)
11. not in use
12. not in use
13. **Zero Standard solution** (2ml, Ready-to-use)
14. **Standard solution 1** (1ml, Ready-to-use) **0.0125 ng/ml**
15. **Standard solution 2** (1ml, Ready-to-use) **0.025 ng/ml**
16. **Standard solution 3** (1ml, Ready-to-use) **0.05 ng/ml**
17. **Standard solution 4** (1ml, Ready-to-use) **0.1 ng/ml**
18. **Standard solution 5** (1ml, Ready-to-use) **0.2 ng/ml**
19. **Standard solution 6** (1ml, Ready-to-use) **0.3 ng/ml**

6. EQUIPMENT AND MATERIALS REQUIRED BUT NOT PROVIDED

- Scales and weighing vessels
- Gloves
- Fume hood
- Homogeniser
- Vortex, mixer
- Centrifuge (4000 x g)
- Automated microtiter plate washer or 8-channel micropipette 100 – 300 µl
- Microtiter plate shaker
- Microtiter plate reader with 450 nm filter
- Micropipettes 0.5 – 10 µl
- Micropipettes, 100 – 1000 µl
- Multipipette with 2.5 ml combi tips
- Sodium acetate

7. PRECAUTIONS

- This kit may contain hazardous substances. For hazard notes please refer to the appropriate safety data sheets (SDS).
- Avoid contact of all biological materials with skin and mucous membranes.
- Do not pipette by mouth.
- Do not eat, drink, smoke, store or prepare foods, or apply cosmetics within the designated work area.
- Do not use components past expiration date and do not use components from different lots.
- Each well is ultimately used as an optical cuvette. Therefore, do not touch the under surface of the wells, prevent damage and dirt.
- All components should be completely dissolved before use. Take special attention to the substrate and rinsing buffer, which crystallize at +4°C.
- Optimal results will be obtained by strict adherence to this protocol. Careful pipetting and washing throughout this procedure are necessary to maintain good precision and accuracy.

R-Biopharm Nederland makes no warranty of any kind, either expressed or implied, except that the materials from which its products are made are of standard quality. There is no warranty of merchantability of this product, or of the fitness of the product for any purpose. R-Biopharm Nederland shall not be liable for any damages, including special or consequential damage, or expense arising directly or indirectly from the use of this product.

8. SAMPLE PREPARATIONS

Mussel, oyster and scallop

- Homogenize 1 g of mollusc meat.
- Add 5 ml of sodium acetate buffer (see chapter 9).
- Mix (vortex) for 1 minute.
- Centrifuge at 3000 x g for 10 minutes at room temperature (20°C - 25°C).
- Save the supernatant.
- Dilute supernatant 1:50 in dilution buffer. (Pipette 10 µl of sample and add 490 µl of dilution buffer (see chapter 9).
- Mix thoroughly
- Use 50 µl per well in the ELISA.

9. PREPARATION OF REAGENTS

Before starting the test, the reagents should be brought up to ambient temperature. Any reagents not used should be put back into storage immediately at +2°C to +8°C.

Microtiter plate

Return unused strips into the resealable zip lock bag with desiccant and store at +2°C to +8°C for use in subsequent assays. Retain also the strip holder.

Rinsing buffer

The rinsing buffer is delivered 20 times concentrated. Prepare dilutions freshly before use. Per strip 20 ml of diluted rinsing buffer is used (1 ml concentrated rinsing buffer + 19 ml distilled water).

Substrate solution

The substrate solution (ready-to-use) precipitates at 4°C. Take care that this vial is at room temperature (keep in the dark) and mix the content before pipetting into the wells.

Dilution buffer

The dilution buffer is 10 times concentrated. Before dilution (10 ml buffer + 90 ml distilled water) the concentrated buffer should be at room temperature and thoroughly mixed. Concentrated buffer can show precipitates of the contents. Mix well before dilution with distilled water.

Saxitoxin Conjugate

Reconstitute the vial of lyophilised conjugate with 4 ml of dilution buffer, mix thoroughly and keep in the dark until use. The reconstituted compound can be stored in a refrigerator (2°C to 8°C) for maximally one week. For longer storage make aliquots and freeze at – 20°C.

Saxitoxin antibody

The saxitoxin antibody is provided in concentrated form. Spin down the antibody in the vial by a short centrifugation (1 minute, 1000 x g). The antibody is diluted 100x in the dilution buffer (e.g. 5 µl antibody + 495 µl of dilution buffer). Per 2x8 wells 400 µl of diluted antibody is required. Store the unused antibody immediately in the dark at +2°C to +8°C.

Sodium acetate buffer (0.2M)

3.28 g anhydrous sodium acetate in 200 ml water (pH 5 ± 0.1 with 1M HCl or 1M NaOH)

10. ASSAY PROCEDURE

Rinsing protocol

In ELISA's, between each immunological incubation step, un-bound components have to be removed efficiently. This is reached by appropriate rinsing. It should be clear that each rinsing procedure must be carried out with care to guarantee good inter- and intra-assay results. Basically, manual rinsing or rinsing with automatic plate wash equipment can be done as follows:

Manual rinsing

1. Empty the contents of each well by turning the microtiter plate upside down followed by a firm short vertical movement.
2. Fill all the wells to the rims (300 µl) with rinsing solution.
3. This rinsing cycle (1 and 2) should be carried out for at least 3 times.
4. Turn the plate upside down and empty the wells by a firm short vertical movement.
5. Place the inverted plate on absorbent paper towels and tap the plate firmly to remove residual washing solution from the wells.
6. Take care that none of the wells dry out before the next reagent is dispensed.

Rinsing with automatic microtiter plate washing equipment

When using automatic plate wash equipment, check that all wells can be aspirated completely, that the rinsing solution is correctly dispensed reaching the rim of each well during each rinsing cycle. The washer should be programmed to execute at least three rinsing cycles.

Assay Protocol

1. Prepare samples according to Chapter 8 and prepare reagents according to Chapter 9.
2. Pipette 100 µl of zero standard in duplicate (well H1, H2).
Pipette 50 µl of zero standard in duplicate (well A1, A2).
Pipette 50 µl of each standard dilution in duplicate (well B1,2 to G1,2, i.e. 0.0125, 0.025, 0.05, 0.1, 0.2 and 0.3 ng/ml).
Pipette 50 µl of each sample solution in duplicate into the remaining wells of the microtiter plate.
3. Add 25 µl of conjugate (HRP) to all wells, except wells H1 and H2.
4. Add 25 µl antibody to all wells, except wells H1 and H2.
5. Seal the microtiter plate and shake the plate for 1 minute.
6. Incubate for 30 minutes at room temperature (20°C – 25°C).

7. Discard the solution from the microtiter plate and wash 3 times with rinsing buffer.
8. Pipette 100 μ l of substrate solution into each well. Mix thoroughly and incubate for 15 min. at room temperature in the dark (20°C - 25°C).
9. Add 100 μ l stop solution to each well.
10. Read the absorbance values immediately at 450 nm.

11. INTERPRETATION OF RESULTS

In general, the O.D. values of the samples, that are measured in this ELISA test, are converted into corresponding concentrations in the following way:

The mean optical density (O.D.) value of the blank wells is subtracted from the individual O.D. of the wells containing the standards and the samples. Then these O.D. values of the standards and the samples (mean values of the duplicates) are divided by the mean O.D. value of the zero standard and multiplied by 100. The zero standard is thus made equal to 100% (maximal absorbance) and the other O.D. values are quoted in percentages of the maximal absorbance. The values calculated for the standards are entered in a semi-logarithmic plot against the concentration of the analyte.

Alternative for calibration curve:

The values of absorption (logit) calculation of the standards are plotted on Y-axis versus the analyte equivalent concentration on a logarithmic X-axis.

As customer-friendly options R-Biopharm offers 2 software programs that can be used for the interpretation of the ELISA test:

- RIDASOFT®Win.NET Food & Feed (art. nr. Z9996FF). In this software program all R-Biopharm ELISA tests (including EuroProxima) are preprogrammed in the database. A list of compatible readers is available on request.
- Simplefit. This Excel based program is developed to calculate the results of all EuroProxima tests.

The calculated saxitoxin equivalents have to be multiplied by 300 to obtain the saxitoxin equivalents in the shellfish tissue.

12. LITERATURE

M. Dubois, L. Demoulin, C. Charlier, G. Singh, S.B. Godefroy, K. Campbell, C.T. Elliott, and Ph. Delahaut (2010). Development of ELISA's for detecting domoic acid, okadaic acid, and saxitoxin and their applicability for the detection of marine toxins in samples collected in Belgium. *Food Additives Contam.* **27(6)**: 859-868.

European Commission 2004. Commission regulation 853/2004/EC of 29 April 2004 laying down specific hygiene rules for food of animal origin. *Off. J. Eur. Comm.* **L226**: 22-82.

K, Campbell, A-C. Huet, C. Charlier, C. Higgins, P. Delahaut, and C.T. Elliott (2009). Comparison of ELISA and SPR biosensor technology for the detection of paralytic shellfish poisoning toxins. *J. Chromatogr.B* **877**: 4079-4089.

13. ORDERING INFORMATION

For ordering the saxitoxin ELISA kit please use cat. Code 5191SAXI.

14. REVISION HISTORY

Chapter 8, addition of explanation to the sample preparation.

Chapter 9, saxitoxin antibody, has been corrected and addition to the preparation of the Sodium Acetate buffer.

Chapter 10, ready-to-use peroxide/TMB changed to substrate solution.

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