



CERTIFICATION

AOAC® Performance TestedSM

Certificate No.

022102

The AOAC Research Institute hereby certifies the method known as:

SureFast® SARS-CoV-2 PLUS Test

manufactured by
Congen Biotechnologie GmbH
Robert-Roessle-Straße 10
13125 Berlin,
Germany

distributed by

R-Biopharm AG	R-Biopharm Inc.
An der neuen	870 Vossbrink Drive
Bergstraße 17	Washington, MO
64297 Darmstadt	63090 USA
Germany	

This method has been evaluated in the AOAC® *Performance Tested MethodsSM* Program and found to perform as stated by the manufacturer contingent to the comments contained in the manuscript. This certificate means that an AOAC® Certification Mark License Agreement has been executed which authorizes the manufacturer to display the AOAC *Performance TestedSM* certification mark along with the statement - "THIS METHOD'S PERFORMANCE WAS REVIEWED BY AOAC RESEARCH INSTITUTE AND WAS FOUND TO PERFORM TO THE MANUFACTURER'S SPECIFICATIONS" - on the above-mentioned method for a period of one calendar year from the date of this certificate (January 27, 2022 – December 31, 2022). Renewal may be granted at the end of one year under the rules stated in the licensing agreement.

A handwritten signature in black ink that reads 'Scott Coates'.

Scott Coates, Senior Director
Signature for AOAC Research Institute

January 27, 2022

Date

METHOD AUTHORS

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SUBMITTING COMPANIES

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MANUFACTURING COMPANY

Congen Biotechnologie GmbH
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13125 Berlin,
Germany

METHOD NAME

SureFast® SARS-CoV-2 PLUS Test

CATALOG NUMBERS

SureFast® SARS-CoV-2 PLUS F7110; SureFast® Prep F1051; SureFast® PCR 7710

INDEPENDENT LABORATORY FOR R-BIOPHARM

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APPLICABILITY OF METHOD

Analytes – SARS-CoV-2 virus

Matrixes – Stainless steel surface (2" by 2" swab)

Performance claims - Performance comparable to the U.S. Centers for Disease Control and Prevention 2019-Novel Coronavirus Real-Time RT-PCR Diagnostic Panel, Revision 04 (2).

REFERENCE METHOD

Centers for Disease Control and Prevention (2020). *CDC 2019-Novel Coronavirus (2019-nCoV) Real-Time RT-PCR Diagnostic Panel. Revision 5. (2)*

ORIGINAL CERTIFICATION DATE

February 10, 2021

CERTIFICATION RENEWAL RECORD

Renewed annually through December 2022.

METHOD MODIFICATION RECORD

1. September 2021

SUMMARY OF MODIFICATION

1. Granted PTM status from ERV.

Under this AOAC® *Performance Tested*SM License Number, 022102 this method is distributed by:
R-Biopharm-AG

Under this AOAC® *Performance Tested*SM License Number, 022102 this method is distributed as:
SureFast® Salmonella ONE

PRINCIPLE OF THE METHOD (1)

The SureFast® SARS-CoV-2 PLUS is a real-time RT-PCR for the direct, qualitative detection of intact novel coronavirus (SARS-CoV-2) RNA from stainless steel swab samples. Each reaction contains an Internal Control RNA (ICR, consisting of MS2-bacteriophage) as an internal control of sample preparation procedure and to monitor possible PCR-inhibition. The RT-qPCR assay can be performed with commonly used real-time PCR instruments, equipped for detection of two fluorescence emissions at the channels FAM and VIC/HEX simultaneously.

DISCUSSION OF THE VALIDATION STUDY (1)

Results from the POD analysis demonstrated that the the SureFast SARS-CoV-2 RT-PCR better at detecting low concentrations (2×10^3 GU/ 2" x 2" test surface) of deposited SARS-CoV-2 on a stainless steel surface compared to the CDC reference method when using the same swabbing sample preparation and swabbing procedure for the both the RT-qPCR primers and probes of the candidate method and the reference method.

The *in silico* analysis of the primers and probes utilized in the SureFast SARS-CoV-2 RT-PCR test method are specific and sensitive enough (99.99% binding of the oligomer and the target binding region) to detect low levels of SARS-CoV-2 without exhibiting false negatives when compared to the CDC reference method. The high level of specificity could be due to the novel single target assay (E gene) requirement of the SureFast SARS-CoV-2 RT-PCR test method in comparison to the double-target assay (N1 and N2 SARS-CoV-2 gene targets) of the CDC reference method. Competition in amplification efficiency between two targets and/or RNA degradation on surfaces may contribute to a single target assay readily detecting one target over a double-target assay. Another possibility for the SureFast SARS-CoV-2 PLUS RT-PCR test method providing better results than the CDC reference method may be due to the difference in swabs used. The swab in the SureFast method may have better recovery of the virus from the stainless steel surface; since there is no prescribed swabbing method in the CDC reference method it is unknown what role the swab material plays in virus recovery.

Inclusivity, Exclusivity and Background Organism Summary (1)

In Silico Analysis	
Inclusivity	
15,764 unique SARS-CoV-2 strain accessions ^a	
Exclusivity	
Human coronavirus (229E, OC43, NL63, HKU1), SARS-coronavirus, MERS-coronavirus, Porcine delta coronavirus	
<u>Background Organisms</u>	
Viruses:	Bovine coronavirus, Human respirovirus 3, Enterovirus, Infectious bronchitis virus, Enterovirus D68, Human adenovirus 1, Human alphaherpesvirus 3, Human bocavirus, Human metapneumovirus, Human orthorubulavirus 2, Human orthorubulavirus 4, Human respirovirus 1, Influenza A H7N9 subtype, Influenza A virus, Influenza A H1N1, Influenza B virus, Norovirus, Respiratory syncytial virus, Simplexvirus, Transmissible gastroenteritis virus
Bacteria and Fungi:	[<i>Candida</i>] <i>glabrata</i> , <i>Acinetobacter baumannii</i> , <i>Acinetobacter baylyi</i> , <i>Acinetobacter bereziniae</i> , <i>Acinetobacter calcoaceticus</i> , <i>Acinetobacter chinensis</i> , <i>Acinetobacter cumulans</i> , <i>Acinetobacter defluvii</i> , <i>Acinetobacter disperses</i> , <i>Acinetobacter equi</i> , <i>Acinetobacter guillouiae</i> , <i>Acinetobacter haemolyticus</i> , <i>Acinetobacter junii</i> , <i>Acinetobacter lactucae</i> , <i>Acinetobacter lanii</i> , <i>Acinetobacter larvae</i> , <i>Acinetobacter nosocomialis</i> , <i>Acinetobacter phage ZZ1</i> , <i>Acinetobacter pittii</i> , <i>Acinetobacter schindleri</i> , <i>Acinetobacter seifertii</i> , <i>Acinetobacter shaoyimingii</i> , <i>Acinetobacter wanghuai</i> , <i>Bacillus cereus</i> , <i>Bacillus thuringiensis</i> , <i>Bordetella pertussis</i> , <i>Candida albicans</i> , <i>Chlamydia pneumoniae</i> , <i>Clostridioides difficile</i> , <i>Enterococcus casseliflavus</i> , <i>Enterococcus cecorum</i> , <i>Enterococcus faecium</i> , <i>Enterococcus hirae</i> , <i>Enterococcus lactis</i> , <i>Enterococcus mundtii</i> , <i>Enterococcus rotai</i> , <i>Enterococcus saigonensis</i> , <i>Enterococcus thailandicus</i> , <i>Enterococcus wangshanyuanii</i> , <i>Escherichia coli</i> O157:H7 str. Sakai, <i>Escherichia coli</i> str. K-12 substr. MG1655, <i>Haemophilus influenzae</i> , <i>Klebsiella pneumoniae</i> , <i>Legionella pneumophila</i> , <i>Listeria monocytogenes</i> , <i>Mycobacterium tuberculosis</i> , <i>Mycoplasma pneumoniae</i> , <i>Pneumocystis jirovecii</i> MT seq, <i>Pseudomonas aeruginosa</i> , <i>Staphylococcus aureus</i> , <i>Staphylococcus epidermidis</i> , <i>Streptococcus pneumoniae</i> , <i>Streptococcus pyogenes</i> , <i>Streptococcus salivarius</i>
Fungi and Eukaryotes:	<i>Homo sapiens aedes aegypti</i> , <i>Aedes albopictus</i> , <i>Dermatophagoides pteronyssinus</i> , <i>Musa domestica</i> , <i>Drosophila</i> , <i>Chlorocebus sabaues</i>

^a Accessions acquired from the Global Initiative on Sharing Avian Influenza Data (GISAID) database from December 2019 to 26 June 2020.

Table 14. Stainless Steel Candidate vs. Reference Method – POD Results (1)

Matrix	Strain	GU/Test Area ^a	N ^b	Candidate SureFast® SARS-CoV-2			Reference			dPOD _c ^f	95% CI ^g
				x ^c	POD _c ^d	95% CI	X	POD _R ^e	95% CI		
Stainless Steel (2" x 2")	SARS-CoV-2 BEI NR-52281	0	5	0	0.00	0.00, 0.43	0	0.00	0.00, 0.43	0.00	-0.43, 0.43
		2.0 x 10 ³	20	20	1.00	0.84, 1.00	11	0.55	0.34, 0.74	0.55	0.20, 0.66
		2.0 x 10 ⁴	5	5	1.00	0.57, 1.00	5	1.00	0.57, 1.00	0.00	-0.43, 0.43

^aGU/Test Area = Results of the GU/Test area were determined by plating the inoculum for each matrix in triplicate

^bN = Number of test portions

^cx = Number of positive test portions

^dPOD_c = Candidate method confirmed positive outcomes divided by the total number of trials

^ePOD_R = Reference method confirmed positive outcomes divided by the total number of trials

^fdPOD_c = Difference between the confirmed candidate method result and reference method confirmed result POD values

^g95% CI = If the confidence interval of a dPOD does not contain zero, then the difference is statistically significant at the 5% level

REFERENCES CITED

- Lacorn, M., Mehl, M., Knoll, C., and Meinhardt, P., Validation of the SureFast® SARS-CoV-2 PLUS Test Method for the Detection of SARS-CoV-2 Virus on Stainless Steel Surfaces, AOAC® Performance TestedSM Emergency Response Validation certification number 022102.
- Centers for Disease Control and Prevention (2020). CDC 2019-Novel Coronavirus (2019-nCoV) Real-Time RT-PCR Diagnostic Panel. Revision 5. 07/13/2020. <https://www.fda.gov/media/134922/download> (Accessed October 2020).